

## Gene Expression Profiling as a Tool for Crop Improvement in Horticultural Biotechnology

Müjgan Güney<sup>1</sup>

Servet Aras<sup>2</sup>

Gökçe Aydöner Çoban<sup>3</sup>

### Abstract

Gene expression profiling has become known as a crucial component of horticulture biotechnology, providing essential insights into the genetic and molecular pathways that influence crop yield, quality, and stress resilience. This overview emphasizes the ideas, applications, and integration of gene expression profiling to improve horticulture crops. High-throughput methodologies, including RNA-Seq, microarray analysis, and qRT-PCR, facilitate the discovery of differentially expressed genes associated with critical agronomic properties like as stress tolerance, fruit quality, and disease resistance. Progress in comprehending microRNA regulation, quantitative trait loci (QTL) mapping, and the discovery of molecular markers has expedited marker-assisted selection and genome editing techniques, including CRISPR/Cas9. The integration of transcriptomic data with other omics technologies—metabolomics, proteomics, and genomics—has enabled a systems-level comprehension of numerous plant processes, including secondary metabolite production and postharvest physiology. Although difficulties are associated with data complexity, elevated expenses, and restricted genetic resources in non-model species, novel methods such as single-cell RNA sequencing and artificial intelligence are improving analytical accuracy. Gene expression profiling ultimately connects molecular insights with applied breeding, facilitating the creation of better, climate-resilient, and nutritionally enhanced horticultural cultivars vital for sustainable agriculture.

- 1 Associate Professor, Yozgat Bozok University, mujgan.guney@yobu.edu.tr, ORCID: 0000-0001-5491-1430
- 2 Associate Professor, Yozgat Bozok University, servet.aras@yobu.edu.tr, ORCID: 0000-0002-0851-8803
- 3 Assistant Professor, Yozgat Bozok University, gokce.aydoner@yobu.edu.tr, ORCID:0000-0002-0347-6552

## 1. Introduction

Gene expression profiling has become an essential tool in horticultural biotechnology, facilitating crop improvement through the elucidation of genetic factors that determine plant traits and responses to environmental stimuli. This approach enables researchers to unravel the complex regulatory networks governing important characteristics such as stress tolerance, growth, and development in various horticultural species.

One of the primary benefits of gene expression profiling is its ability to identify differentially expressed genes (DEGs) under specific conditions. For instance, studies in banana (*Musa acuminata* L.) have identified genes related to stress response and hormone signaling pathways that can be targeted to enhance drought and cold tolerance, thereby improving crop productivity in water-limited environments and cold conditions (Xu et al., 2023). Similar results have been reported in tomato, where the transcription factor SIHY5 was found to regulate fruit ripening through complicated gene interactions linked to environmental signals (Wang et al., 2021). These insights are crucial for developing cultivars that can thrive under unfavorable conditions, hence enhancing horticultural practices (Guney et al, 2024a).

Moreover, gene expression analysis provides a foundation for the application of advanced genomic tools such as CRISPR/Cas9 technology. This technology allows for targeted editing of gene expressions and regulatory elements, thereby it makes the improving crops with desirable traits (Li et al., 2020; Ku & Ha, 2020). By understanding the specific genes involved in phenotypic variation, researchers can utilize integration of CRISPR methodologies and gene expression data to introduce new cultivars with improved nutritional profiles or increased resistance to pests and diseases (Guney et al, 2024b).

In addition to stress responses, gene expression profiling is also instrumental in addressing horticultural challenges such as fruit quality and yield. Research on peach (*Prunus persica*) has highlighted the crucial role of genomic analyses in elucidating the genetic basis of domestication traits, which can inform breeding strategies aimed at improving fruit characteristics such as size and sweetness (Cao et al., 2019; Bie et al., 2023). Notably, the production of antioxidant compounds, essential for maintaining fruit quality during storage, has also been linked to specific gene expressions influenced by various environmental factors, underscoring the potential for gene profiling in enhancing postharvest management practices (Özyalın, 2023; Guney et al., 2022; Ergun, 2021).

Moreover, numerous tools like dynamic expression atlases have been established for promoting functional genomics research in horticulture crops. For example, FEAtl is a web-based platform that catalogs gene expressions in tropical and subtropical fruit crops, aiding in the identification of genetic factors related to production challenges (Roy et al., 2024). Such databases enable researchers to perform large-scale analyses and promote collaboration across the field of horticultural biotechnology, emphasizing the importance of gene expression profiling in sustaining future crop improvement initiatives.

In conclusion, gene expression profiling stands as a basis of horticultural biotechnology, enabling detailed insights into the genetic underpinnings of plant traits and their responses to environmental conditions. By advancing our understanding of gene regulatory mechanisms and facilitating the application of biotechnological tools, gene expression analysis paves the way for sustainable agricultural practices and the enhancement of crop productivity.

## **2. Principles of Gene Expression Profiling**

Gene expression profiling is a technique employed to assess the activity of numerous genes simultaneously, hence generating an extensive overview of cellular function. In horticultural biotechnology, this technique is vital for understanding the genetic basis of traits, enabling the enhancement of crop characteristics such as yield, disease resistance, and abiotic stress tolerance. By examining gene expression patterns, researchers can identify important molecular players involved in various physiological processes that directly affect plant performance.

A foundational aspect of gene expression profiling involves the use of high-throughput sequencing technologies, such as RNA-seq, which allows for the comprehensive analysis of transcriptomes in horticultural crops. RNA-seq has provided significant insights into the temporal and Tissue-specific expression of genes linked to important traits in a variety of species. For example, studies in avocado have identified transcripts associated with phenological events that correlate with seasonal changes, highlighting the relevance of gene expression data in predicting plant behavior and adaptability (Ahsan et al., 2023).

Understanding the molecular regulation of microRNAs (miRNAs) is another critical dimension of gene expression profiling in horticultural plants. miRNAs are small non-coding RNAs that modulate gene expression and have been implicated in plants' stress responses. Recent research has shown that specific miRNAs can enhance disease resistance in various crops,

illustrating the potential of miRNA profiling as a strategic tool in plant breeding programs (Zhang et al., 2023). The identification and validation of these microRNAs may lead to the development of crops that are better equipped to handle biotic stresses, thereby enhancing food production against pest invasions.

Another principle inherent in gene expression profiling involves the genetic mapping of quantitative trait loci (QTLs) to better understand heritable traits in crops. QTL analysis has successfully identified genetic markers associated with important horticultural traits such as fruit size, quality, and resistance to environmental stressors in crops like melon Zhao et al. (2023) and lettuce (Macías-González et al., 2021). Integrating gene expression data with QTL mapping enhances the precision of marker-assisted selection (MAS) strategies, thus accelerating the breeding process for desirable traits (Guney et al., 2018).

The development of robust molecular markers for breeding also relies on gene expression profiling. For instance, studies have resulted in the identification of functional genetic markers that can be utilized in the selection of clonal rootstocks for apple, demonstrating how genetic analyses can lead to improved horticultural practices (Shamshin et al., 2023). Marker-assisted breeding, boosted by gene expression data, facilitates the more efficient selection of plants with desirable traits, overcoming the limitations of traditional breeding techniques that often involve prolonged trial-and-error processes (Guney et al, 2021).

Methodically, gene expression profiling employs various bioinformatics tools to analyze expression data and identify significant differentially expressed genes (DEGs). These DEGs serve as potential targets for functional validation and further exploration regarding their roles in plant biology. For example, transcriptome analyses have provided insights into the genetic and physiological mechanisms underpinning abiotic stress tolerance in crops like Chinese cabbage Su et al. (2019) and litchi (Zhou et al., 2022). Such studies connect environmental responses to genetic expression pathways, offering a clearer understanding of resilience traits in horticultural species.

In conclusion, gene expression profiling serves as a dynamic and integral component of horticultural biotechnology, significantly enhancing our understanding of plant gene function and regulation. The interplay between high-throughput sequencing technologies, microRNA functionality, genetic mapping, and molecular marker development establishes a framework for advancing crop improvement strategies. With ongoing research, the

application of gene expression profiling will continue to empower scientists and breeders in their efforts to optimize and innovate horticultural practices.

### 3. Applications in Horticultural Crop Improvement

#### 3.1. Enhancing Stress Tolerance

Improving stress tolerance in horticulture crops is a vital goal in biotechnological interventions, particularly due to climate change and rising environmental stressors. Stress tolerance is crucial for maintaining crop productivity and quality under adverse conditions such as drought, salinity, extreme temperatures, and diseases. Advances in gene expression profiling and molecular biotechnology have provided significant insights into the mechanisms underlying plant stress responses, allowing for the development of varieties that can withstand these challenges.

One of the promising strategies for enhancing stress tolerance involves the application of phytohormones such as melatonin, tryptophan, salicylic acid, and abscisic acid. Melatonin, in particular, has demonstrated multifaceted roles in bolstering drought stress tolerance in crops like tomatoes by improving root architecture, enhancing photosynthesis, and fortifying antioxidant defense systems (Altaf et al., 2022; Kumari et al., 2023, Balci et al., 2023). It modulates stress responses through the regulation of redox homeostasis and increases the activities of various antioxidant enzymes, effectively scavenging reactive oxygen species (ROS) produced during stress (Altaf et al., 2022; Tiwari et al., 2021). Research has shown that melatonin application can also increase the expression of heat shock proteins, which are crucial in protecting cellular functions during temperature fluctuations (Zhang et al., 2017).

MicroRNAs (miRNAs) are another area of focus, as they play a vital role in mediating stress responses in plants. For instance, studies on bananas have elucidated the involvement of temperature-responsive miRNAs that target transcription factors associated with auxin signaling and redox homeostasis, revealing their potential for use in genetic improvement programs aimed at enhancing abiotic stress resistance (Zhu et al., 2019). Understanding the regulatory networks governed by miRNAs can offer avenues for precise genetic modifications that boost crop resilience.

Biotechnological applications also utilize genomic approaches, such as Genome-Wide Association Studies (GWAS) and marker-assisted selection (MAS), to identify key genes and quantitative trait loci (QTLs) responsible for stress tolerance. Research on apple has highlighted the potential of

integrating genomic methods with traditional breeding techniques to enhance abiotic stress tolerance (Dutta et al., 2022). Similarly, studies involving transgenic approaches have successfully discussed salt tolerance to various horticultural crops by transferring genes associated with stress responses, including those coding for glycine-betaine synthesis and regulatory transcription factors (Parmar et al., 2017; Wang et al., 2016). This genetic engineering can be pivotal in creating varieties with enhanced resistance to abiotic stresses such as salinity and drought.

Additionally, studies have documented the role of brassinosteroids in improving resilience against abiotic stressors. Brassinosteroids regulate various physiological and biochemical pathways that bolster stress tolerance, consequently supporting overall plant health under adverse conditions (Zhang et al., 2023). Their integration into crop management strategies can significantly improve the yield stability of horticultural crops subjected to environmental stress.

Furthermore, abiotic stress management in horticultural crops also emphasizes the importance of biostimulants, which are substances that enhance plant growth and resistance under stress conditions. These can include natural extracts such as seaweed, which is rich in phytohormones and essential nutrients, promoting growth and resilience in crops (Tejasree et al., 2024). Biostimulants contribute to stress mitigation by enhancing the plant's nutrient uptake and assimilation capabilities. Research has demonstrated that interventions like hydrogen-rich water and nitric oxide sprays can improve the stress tolerance of crops. These therapies are recognized for activating signaling pathways that regulate gene expression associated with stress responses (Sun et al., 2021; Gong et al., 2017). The control of ethylene levels has been shown to enhance cold tolerance in apple crops, highlighting the critical role of hormone regulation in stress adaptation (Wang et al., 2021).

### **3.2 Improving Fruit Quality and Postharvest Traits**

The enhancement of fruit quality and postharvest traits is essential for maximizing the marketability and consumer acceptance of horticultural crops. A broad range of techniques, including the application of growth regulators, biostimulants, and novel postharvest treatments, have been systematically employed to enhance attributes of fruits such as flavor, texture, color, nutritional content, and shelf life (Ergun and Bozkurt, 2020;).

One notable approach in improving fruit quality involves the application of brassinosteroids, which are plant hormones that play a pivotal role in

various physiological processes. The gene *SICYP90B3* has been identified as a significant regulator of tomato fruit quality, influencing ripening processes, softening, and the accumulation of flavor volatiles, soluble sugars, and carotenoids. Manipulating the expression of *SICYP90B3* can lead to enhanced fruit quality attributes, including improved flavor and visual appeal, thereby increasing consumer acceptance (Hu et al., 2022).

Bioactive compounds, particularly flavonoids, are crucial for both fruit quality and resistance to postharvest diseases. The MADS-box protein *SITAGL1* in tomatoes regulates ripening-associated flavonoid biosynthesis, contributing to both enhanced fruit quality and resistance to pathogens like *Botrytis cinerea* (Wang et al., 2023). Similarly, the transcription factor *SlbHLH95* has been implicated in the regulation of flavonoid metabolism, highlighting the potential for targeted genetic strategies to improve resistance to postharvest diseases while enhancing nutritional quality (Su et al., 2025). This underscores the importance of understanding the genetic and molecular mechanisms that underpin fruit quality traits.

Foliar application of compounds such as forchlorfenuron (CPPU) has been shown to significantly affect the biochemical indices and overall quality of crops. A study demonstrated that CPPU enhanced sugar transport from leaves to fruit, thereby improving fruit retention and quality in fig trees (Abdel-Azeem et al., 2023). Such chemical treatments can effectively enrich fruit quality by addressing physiological aspects related to sugar accumulation and metabolic processes.

Beyond chemical applications, the role of nitric oxide (NO) during postharvest management is also worthy of attention. Research indicates that exogenous applications of NO not only delay fruit senescence but also enhance fruit quality attributes, increase antioxidant activity, and mitigate chilling injuries during storage (Wei et al., 2024). The metabolic pathways stimulated by nitric oxide enhance fruit quality preservation, offering strong strategies to combat postharvest decline.

Innovative techniques for fruit preservation, including the use of ethylene scavengers and protective coatings, significantly contribute to maintaining postharvest fruit quality. The combination of potassium permanganate with ultraviolet light has been shown to preserve sensory attributes of peaches, enhancing both taste and aroma which are important factors that influence consumer preferences (Alonso-Salinas et al., 2023). Techniques that focus on preserving the initial aroma and sensory quality of fruits extend their market potential and consumer appeal.



The application of bagging techniques in fruit production has emerged as an effective method for enhancing the external appearance and quality of fruits. Preharvest bagging has been associated with improved coloration, size uniformity, and reduced rates of fungal infections (Buthelezi et al., 2023). Moreover, bagging not only protects fruits from environmental stressors but also promotes secondary metabolite biosynthesis, thereby enriching the nutritional composition and flavor profiles of fruits. In addition to bagging, the innovation of using biodegradable films for fruit preservation has gained traction. Recent developments in green bioactive films have shown promise in protecting fruit quality by minimizing exposure to pathogens and controlling respiration rates, thus prolonging shelf life (Ahmed et al., 2022,). The incorporation of natural extracts into these films provides added health benefits, enhancing the marketability of fresh produce. Therefore, Transcriptomic analyses of bagged fruits have demonstrated notable differential expression in genes associated with secondary metabolism, especially those implicated in flavonoid biosynthesis, hormonal regulation, and defense mechanisms, suggesting that bagging modifies fruit microenvironments and initiates molecular reprogramming.

Non-destructive testing technologies, such as near-infrared spectroscopy and advanced image analysis, are instrumental in assessing fruit quality parameters without damaging the product. These techniques offer rapid and precise evaluations of attributes like fruit maturity, sweetness content, and external appearance, enabling improved decision-making in postharvest processing and marketing (Kusumiyati et al., 2019; Kim et al., 2024). Understanding and applying these analytical approaches can significantly enhance the efficiency of the fruit supply chain, ensuring that only high-quality produce reaches consumers. Although non-destructive approaches do not directly quantify gene expression, they record phenotypic characteristics such as color, hardness, and biochemical composition—that are ultimately governed by underlying gene activity (Hernández-Hierro et al., 2022).

### **3.3. Disease Resistance**

The enhancement of disease resistance in horticultural crops is a critical focus of research aimed at ensuring sustainable agricultural practices and enhancing food security. Various approaches have been developed to improve disease resistance, utilizing both traditional breeding techniques and modern biotechnological methodologies.

One of the promising strategies involves the application of biostimulants like melatonin, which has been shown to enhance resistance against various



pathogens. Studies reveal that exogenous melatonin can improve the resistance of apple (*Malus domestica*) to the fungal pathogen *Marssonina*, which causes significant foliar damage. This protective role is attributed to the ability of melatonin to modulate oxidative stress responses, leading to enhanced reactive oxygen species (ROS) generation that can inhibit pathogen growth (Yin et al., 2013). Melatonin also plays a crucial role in enhancing the general stress tolerance of horticultural crops, contributing to their resilience against biotic stress (Hao et al., 2025).

Natural plant extracts have gained attention as a means of improving disease resistance in horticultural crops. For instance, research has demonstrated that specific extracts can effectively control fungal pathogens, thereby improving productivity in crops like zucchini (*Cucurbita pepo*). Flavonoids, which are abundant in these extracts, exhibit antimicrobial properties and can play dual roles as defense signals as well as UV protectants (Hassan et al., 2021). The exploration of these natural compounds as biopesticides provides an environmentally friendly alternative to synthetic pesticides, which are often associated with chemical residues and ecological concerns.

Chitosan, a biopolymer derived from crustacean shells, has been reported to induce systemic resistance against pathogens like *Fusarium oxysporum* in tomato. The mechanism involves the upregulation of specific pathogenesis-related (PR) proteins and antioxidants, which enhance the plant's defensive abilities. Chitosan acts as both a physical barrier and a biochemical stimulus, fostering an innate immune response that can be utilized in integrated disease management systems (Carmona et al., 2021; Güney et al., 2024c).

A contemporary advancement in enhancing disease resistance involves the use of genome editing technologies, particularly CRISPR/Cas9. This powerful tool allows for precise modifications of plant genomes to improve resistance against specific pathogens. For instance, transgenic crops expressing resistance genes or gene-editing constructs can improve resilience to diseases caused by fungi, bacteria, and viruses (Tyagi et al., 2021). Such genetic engineering techniques enable the development of varieties that are tailored for enhanced disease resistance while maintaining desirable agronomic traits.

The NPR1 gene, known for its role in initiating systemic acquired resistance, has been successfully integrated into various horticultural crops to boost their defense mechanisms against pathogens. Studies show that overexpression of NPR1 enhances the disease resistance responses of plants, particularly against pathogens such as *Xanthomonas citri*, responsible for citrus canker (Zhang et al., 2010). This approach illustrates how biotechnological

strategies can lead to significant improvements in the health and survival of crops under pathogen pressure.

The use of light quality manipulation has also emerged as an innovative method to enhance disease resistance. Research indicates that exposure to specific wavelengths of light, particularly the red and far-red spectrum, can improve the resistance of cucumber seedlings to powdery mildew. This physiological defense mechanism is linked to enhanced antioxidant activity and the accumulation of signaling molecules involved in stress responses (Shibuya et al., 2011).

Fungal elicitors, such as those derived from pathogens, have shown potential in activating defense responses in crops. The application of elicitors can stimulate the innate immunity of plants, conferring resistance to various pathogens, including *Botrytis cinerea*, which causes significant losses in multiple horticultural crops (Perato et al., 2020; Li & Cheng, 2023). The mechanisms underlying these defense responses involve complex signaling pathways that can be harnessed for developing resilient crop varieties.

Moreover, even traditional approaches such as grafting have proven effective in improving disease resistance. Grafting horticultural plants onto resistant rootstocks has been widely practiced to manage soil-borne diseases and enhance overall plant health. This technique allows for the combination of a robust root system with a productive scion, significantly enhancing resistance against pathogens and improving yield outcomes (Goldschmidt, 2014).

In conclusion, the quest for improving disease resistance in horticultural crops encompasses a multitude of strategies ranging from the application of natural biostimulants and elicitors to advanced genetic engineering techniques. By integrating these methods, researchers can develop sustainable solutions to combat plant diseases, ensuring the viability and productivity of important horticultural species in an era of increasing agricultural challenges.

### **3.4. Understanding Secondary Metabolism**

The study of secondary metabolism in horticultural plants is crucial as it encompasses the biochemical pathways leading to the synthesis of a wide variety of compounds that play essential roles in plant defense, flavor, aroma, and color. These metabolites greatly influence the nutritional, medicinal, and economic value of horticultural crops. Advances in molecular biology, proteomics, and genomics have provided significant insights into the regulatory mechanisms that govern the production of secondary metabolites.

Phenolic compounds, including flavonoids and phenolic acids, have been recognized for their important roles in plant defense mechanisms and human health (Özyalın and Yaman, 2023). In particular, recent studies have demonstrated that environmental factors such as light exposure and stress can dramatically influence the accumulation of these compounds (Zeng et al., 2011). Instead, studies have shown that manipulation of light quality through LED technology can increase the concentration of beneficial secondary metabolites in various crops (Arif et al., 2024).

Moreover, the interaction between environmental stresses and secondary metabolism has been the subject of considerable research. For instance, exposure to ozone stress in *Brassica campestris* has been shown to significantly impact the growth and secondary metabolite production of plants (Han et al., 2023). Such responses illustrate how secondary metabolism is intricately linked to the adaptive strategies under stress conditions in plants.

In apple, studies have explored the regulation of secondary metabolite biosynthesis by bHLH and MYC transcription factors. Xie et al. (2012) reported that the antagonistic relationship between SmbHLH60 and SmMYC2 plays a crucial role in the regulation of phenolic acids and anthocyanins, two important classes of secondary metabolites known for their health benefits. This regulation highlights the complexity of molecular networks that control secondary metabolite synthesis and indicates potential targets for metabolic engineering to enhance the nutritional value of crops.

Melatonin, a small molecule with significant roles in plant development, has also been involved in secondary metabolism. While the current literature reviews the impact of melatonin on antioxidant responses, there is insufficient evidence linking melatonin directly to enhanced secondary metabolites in postharvest horticultural products (Zhang et al., 2022). Further research is required to clarify these connections.

Additionally, phenylpropanoid metabolism, which is pivotal in producing various secondary metabolites such as flavonoids and lignin, has gained attention for its role in environmental interactions. Cold stress, for instance, has been noted to activate the phenylpropanoid pathway, increasing the synthesis of valuable compounds that enhance plant resilience (Kang et al., 2025). Understanding these metabolic shifts is essential for developing strategies to improve crop productivity and resilience against climate-related challenges.

Compounds like carotenoids are another class of secondary metabolites that are crucial not only for plant health but also for human nutrition. They

contribute to the color of fruits and vegetables and have essential antioxidant properties. Recent advancements highlight the intricate regulation of carotenoid biosynthesis through various transcription factors, thus opening avenues for biofortification efforts aimed at increasing provitamin A content in crops like tomato and watermelon (Zheng et al., 2020; Moreno et al., 2020).

The integration of high-throughput omics technologies, including transcriptomics, proteomics, and metabolomics, has significantly advanced the understanding of secondary metabolism in horticultural crops. These technologies enable researchers to elucidate metabolic pathways, identify regulatory genes, and explore the functional genomics of secondary metabolite production (Xu & Xian-pu, 2025). For example, studies combining metabolomics with transcriptomic data have revealed the mechanisms by which phenolic acids are synthesized in grape varieties, providing insight into potential breeding targets for enhanced phytochemical content (Cheng et al., 2023).

#### **4. Integration with Other Omics Technologies**

The combination of diverse omics methods—such as genomics, transcriptomics, proteomics, and metabolomics has transformed the comprehension of intricate biological processes in horticulture plants. This multi-omics methodology enables researchers to investigate the complex interconnections of genes, proteins, and metabolites, resulting in a comprehensive comprehension of plant development, stress responses, and secondary metabolite synthesis.

One significant application of integrating metabolomics and transcriptomics is the study of anthocyanin biosynthesis in horticultural crops. For instance, research by Juxian et al. (2022) highlighted that the combination of transcriptomic and metabolic data reveals how light conditions influence anthocyanin accumulation in *Brassica* plants. This study illustrated that different light regimes directly impact the expression of genes involved in anthocyanin synthesis, subsequently affecting metabolite levels (Juxian et al., 2022). Such findings underline the importance of light as a regulatory factor and demonstrate how integrating omics data can elucidate the mechanisms governing secondary metabolite production, further enhancing fruit quality.

In blueberries, the dual analysis of small RNA and transcriptomic data has unveiled the regulatory roles of microRNAs during fruit development. In a study, specific miRNAs and their target genes implicated in ripening

processes, emphasizing the significance of small RNAs in modulating developmental pathways that contribute to fruit quality (Hou et al., 2017). This approach showcases how integrating transcriptomic data with small RNA studies provides a more comprehensive understanding of the regulatory networks underpinning fruit ripening.

The metabolic profiling of herbaceous plants, such as *Paeonia lactiflora*, together with transcriptomic data, aids in identifying key genes involved in oil biosynthesis within seeds. Xu et al. (2024) utilized this integrated analysis to reveal the genetic framework contributing to triacylglycerol accumulation, crucial for optimizing oil content in horticultural crops. Such studies exemplify how multi-omics strategies can facilitate targeted breeding studies aimed at enhancing specific agriculturally important traits.

In kiwifruit (*Actinidia chinensis*), an integrative analysis of the metabolome and transcriptome was employed to uncover gene regulatory networks associated with flavor formation. Their findings highlighted the contributions of specific transcription factors in controlling the biosynthesis of flavor compounds, illustrating the potential of omics integration to enhance flavor profile optimization in horticultural crops (Wang et al., 2021).

Additionally, the integration of transcriptomics with metabolomics has been pivotal in understanding flower color differentiation in various plants. Another study discussed how a combined approach revealed the relationship between anthocyanin metabolism and the specific genetic controls that influence petal pigmentation in *Camellia reticulata*. This analysis provides insights into color regulation and aids in selecting traits for ornamental plant breeding (Geng et al., 2022).

The outcome of integrating these omics technologies extends beyond basic research. For example, studies have demonstrated that high-throughput phenotyping methods, used in conjunction with omics analyses, can enhance the efficiency of breeding programs. This integration allows for a better selection of traits based on phenotypic and genotypic data, facilitating the development of new horticultural varieties that meet market demands while maintaining desirable characteristics (Cembrowska-Lech et al., 2023; Zhang et al., 2022).

Furthermore, the application of multi-omics approaches has profound implications for addressing abiotic stresses in horticultural crops. By integrating transcriptomics and metabolomics, researchers can identify physiological and biochemical responses to stressors such as heat, drought,

or cold. For instance, the use of omic strategies to enhance crop resilience to heat stress reported how this integration could reveal the underlying mechanisms by which plants adapt to temperature fluctuations, guiding breeding for climate resilience in horticultural plants (Zhou et al., 2022).

## 5. Challenges and Future Prospects

The application of advanced biotechnology methods in horticulture, including multi-omics, genome editing, and precision agriculture, has demonstrated considerable potential for improving crop yield and sustainability. Nonetheless, numerous problems remain that must be resolved to effectively harness the advantages of these technologies in the horticultural sector. A major challenge facing the horticultural sector relates to the integration of multi-omics data.

While the collective application of genomics, transcriptomics, proteomics, and metabolomics offers a comprehensive understanding of plant biology, the complexity of data integration poses significant problems. Research indicates that discrepancies in data compatibility, quality control issues, and the lack of standardized protocols create barriers to effectively consolidating multi-omics data sets (Cembrowska-Lech et al., 2023). The effective use of artificial intelligence and machine learning may provide solutions for better handling and interpreting large and complex omic data. Future efforts should focus on developing frameworks that enhance data interoperability among different omics technologies, facilitating a holistic understanding of plant phenotypes.

Another critical challenge is the adoption of genome editing technologies, such as CRISPR/Cas9, in the horticultural sector for crop improvement. Despite its potential to generate crops with desirable traits like disease resistance and abiotic stress tolerance, public acceptance and regulatory patterns remain significant obstacles (Bhavaneet al., 2024; Priyanka et al., 2025). Genetic modification techniques face intense analysis and varying regulations across different countries, which can slow down research translation into practical applications in horticulture (Bhavaneet al., 2024). Future research must focus on developing frameworks for transparent communication regarding the safety and benefits of genome-edited crops to bolster public trust.

Moreover, while advances in protected cultivation practices offer opportunities to enhance fruit quality and yield, challenges such as the high cost of initial setup and energy requirements persist (Jain et al., 2023). Achieving sustainability in protected horticulture requires comprehensive

evaluation of resource management techniques and the adoption of renewable energy sources to mitigate operating costs and environmental effects (Jain et al., 2023). Research should concentrate on innovative methodologies that enhance resource utilization efficiency and diminish economic obstacles for producers, particularly smallholder farmers.

Climate change also poses an ongoing challenge for horticultural crop production, affecting yields and fruit quality across various regions (Deori et al., 2024). Horticulture is particularly vulnerable to the changing climate due to its dependence on specific environmental conditions for optimum growth. Future research should prioritize developing climate-resilient varieties through breeding and genetic engineering, as discussed in studies exploring the adaptation of horticultural crops to climate unpredictability (Deori et al., 2024). Additionally, integrating climate-smart agricultural practices into the horticultural sector can help mitigate these challenges and enhance long-term sustainability.

In light of the rapid advancements in technology, another crucial prospect involves the integration of automation and robotics for efficiency in horticulture. While autonomous systems can significantly improve productivity, challenges related to equipment adaptability, crop variability, and the delicate handling of horticultural produce need to be addressed (Sharma et al., 2024). Ongoing research in developing resilient, intelligent automation systems that can adapt to various horticultural conditions will be crucial for optimizing operations and improving labor efficiency.

Furthermore, the lack of skilled horticulturists and researchers to utilize emerging technologies is a barrier to innovation in the horticultural landscape (Pitt, 2021). Educational institutions must adapt curricula to include training in new technologies and sustainable practices. Prospects must emphasize the importance of developing a skilled workforce capable of navigating and utilizing the latest advancements in horticulture.

## **6. Conclusion**

The integration of gene expression profiling with advanced biotechnological tools has revolutionized horticultural science, offering precise and innovative strategies to address key challenges in crop improvement. By enabling the identification of differentially expressed genes (DEGs), regulatory microRNAs, and key transcription factors, gene expression profiling lays a robust foundation for understanding the complex genetic networks that govern traits such as stress tolerance, disease resistance, fruit quality, and postharvest performance.



High-throughput technologies like RNA-seq have empowered researchers to uncover the molecular basis of plant responses to environmental signals, while genome editing tools such as CRISPR/Cas9 have allowed for the precise modification of target genes, accelerating the development of improved cultivars. The regulation of secondary metabolites by transcription factors like MYC and bHLH further highlights how gene expression influences nutritional and commercial traits, with external factors such as melatonin, brassinosteroids, and light quality offering practical means of modulation.

Emerging tools, including non-destructive phenotyping methods and multi-omics approaches (e.g., genomics, transcriptomics, metabolomics), are enhancing the accuracy and efficiency of breeding programs by providing a comprehensive understanding of crop physiology and biochemical composition. Platforms like expression atlases and integrated databases also enable large-scale data analysis and global collaboration, supporting more informed decisions in cultivar development.

The future of horticultural biotechnology relies on an integration of omics technologies, computational tools, and sustainable practices. However, obstacles persist in aligning regulatory frameworks for genome editing, maintaining data compatibility across platforms, mitigating climate change impacts, and enhancing competencies in the advanced technical workforce. Resolving these difficulties necessitates synchronized efforts in research, education, and policy.

Continuous investment in research and innovation enables horticulture biotechnology to make substantial contributions to food security, environmental sustainability, and agricultural resilience. As global demands increase, the significance of cultivating resilient, high-quality horticulture crops capable of thriving in diverse and evolving conditions also rises.

## References

- Abdel-Azeem, M., Beheiry, H., Mohamed, I., Abd-El-Samad, A., Belal, H., Mahmoud, A., ... & Rady, M. (2023). Effect of foliar spraying with forchlorfenuron on late fruit quality and physio-biochemical indices of fig fruits. *Labyrinth Fayoum Journal of Science and Interdisciplinary Studies*, 1(3), 40-46.
- Adegbaju, M., Ajose, T., Adegbaju, I., Omosebi, T., Ajenifujah-Solebo, S., Falana, O., ... & Akinbo, O. (2024). Genetic engineering and genome editing technologies as catalysts for Africa's food security: the case of plant biotechnology in Nigeria. *Frontiers in Genome Editing*, 6. <https://doi.org/10.3389/fgeed.2024.1398813>
- Ahmed, W., Azmat, R., Khojah, E., Ahmed, R., Qayyum, A., Shah, A., ... & Samra, B. (2022). The development of a green innovative bioactive film for industrial applications as a new emerging technology to protect the quality of fruits. *Molecules*, 27(2), 486.
- Ahsan, M., Barbier, F., Hayward, A., Powell, R., Hofman, H., Parfitt, S., ... & Mitter, N. (2023). Molecular cues for phenological events in the flowering cycle in avocado. *Plants*, 12(12), 2304.
- Alonso-Salinas, R., López-Miranda, S., Báidez, A., Pérez-López, A., Noguera-Artiaga, L., Núñez-Delgado, E., ... & Acosta-Motos, J. (2023). Effect of potassium permanganate, ultraviolet radiation and titanium oxide as ethylene scavengers on preservation of postharvest quality and sensory attributes of broccoli stored with tomatoes. *Foods*, 12(12), 2418.
- Altaf, M., Shahid, R., Ren, M., Naz, S., Altaf, M., Khan, L., ... & Ahmad, P. (2022). Melatonin improves drought stress tolerance of tomato by modulating plant growth, root architecture, photosynthesis, and antioxidant defense system. *Antioxidants*, 11(2), 309.
- Arif, A., Budiyo, A., Setiawan, S., Cahyono, T., Sulistiyani, T., Marwati, T., ... & Iswari, K. (2024). Application of red and blue led light on cultivation and postharvest of tomatoes (*Solanum lycopersicum* L.). *Scientifica*, 2024(1).
- Balcı, G., Özyalın, S., & Çoban, G. A. (2023). Dışsal Triptofan Uygulamasının Böğürtlerde Bazı Biyokimyasal ve Meyvelerinin Kalite Kriterleri Üzerine Etkileri. *Bozok Tarım ve Doğa Bilimleri Dergisi*, 2(1), 31-40.
- Berger, L., Stamford, N., Willadino, L., Laranjeira, D., Lima, M., Malheiros, S., ... & Stamford, T. (2016). Cowpea resistance induced against *Fusarium oxysporum* f. sp. tracheiphilum by crustaceous chitosan and by biomass and chitosan obtained from *Cunninghamella elegans*. *Biological Control*, 92, 45-54. <https://doi.org/10.1016/j.biocontrol.2015.09.006>
- Bhavane, K., Krishnamoorthi, A., Rathva, H., Mareguddikar, S., Singh, A., Singh, B., ... & Chittibomma, K. (2024). Advancements in genetic engi-

- neering for enhanced traits in horticulture crops: a comprehensive review. *Journal of Advances in Biology & Biotechnology*, 27(2), 90-110.
- Bie, H., Wang, H., Wang, L., Li, Y., Fang, W., Chen, C., ... & Cao, K. (2023). Mining genes related to single fruit weight of peach (*Prunus persica*) based on WGCNA and GSEA. *Horticulturae*, 9(12), 1335.
- Bigini, V., Camerlengo, F., Botticella, E., Sestili, F., & Savatin, D. (2021). Biotechnological resources to increase disease-resistance by improving plant immunity: a sustainable approach to save cereal crop production. *Plants*, 10(6), 1146.
- Buthelezi, N., Olowoyo, J., & Mafeo, T. (2023). Influence of preharvest bagging on maturity indices and postharvest quality of cherry tomato (*Solanum lycopersicum* var. *cerasiforme*). *Hortscience*, 58(1), 95-104.
- Cao, K., Li, Y., Deng, C., Gardiner, S., Zhu, G., Fang, W., ... & Wang, L. (2019). Comparative population genomics identified genomic regions and candidate genes associated with fruit domestication traits in peach. *Plant Biotechnology Journal*, 17(10), 1954-1970.
- Cembrowska-Lech, D., Krzemińska, A., Miller, T., Nowakowska, A., Adamski, C., Radaczyńska, M., ... & Mikiciuk, M. (2023). An integrated multi-omics and artificial intelligence framework for advance plant phenotyping in horticulture. *Biology*, 12(10), 1298.
- Cheng, G., Wu, D., Guo, R., Li, H., Wei, R., Zhang, J., ... & Zhou, S. (2023). Chromosome-scale genomics, metabolomics, and transcriptomics provide insight into the synthesis and regulation of phenols in *Vitis adenoclada* grapes. *Frontiers in Plant Science*, 14.
- Deori, M., Manasa, S., Kakade, P., Saikanth, D., Ranganna, G., Deshmukh, R., ... & Prasad, L. (2024). A comprehensive review on the impact of climate change on fruit yield and quality in modern horticultural practices. *International Journal of Plant & Soil Science*, 36(1), 177-187.
- Dutta, M., Singh, R. K., & Zinta, G. (2022). Genomic approaches to improve abiotic stress tolerance in apple (*Malus × domestica*). In *Genomic Designing for Abiotic Stress Resistant Fruit Crops* (pp. 1-17). Cham: Springer International Publishing.
- Ergun, Z., & Bozkurt, T. (2020). Determination of fatty acid composition and antioxidant activity of fig seed oil. *International Journal of Agricultural and Natural Sciences*, 13(2), 101-107.
- Ergün, Z. (2021). Seed oil content and fatty acid profiles of endemic *Phoenix theophrasti* Greuter, *Phoenix roebelenii* O'Brien, *Phoenix canariensis* Hort. Ex *Chabaud*, and *Phoenix dactylifera* L. grown in the same locality in Turkey. *Turkish Journal of Agriculture and Forestry*, 45(5), 557-564.
- Geng, F., Nie, R., Yang, N., Cai, L., Hu, Y., Chen, S., ... & Chen, L. (2022). Integrated transcriptome and metabolome profiling of *camellia reticulata*

- reveal mechanisms of flower color differentiation. *Frontiers in Genetics*, 13.
- Goldschmidt, E. (2014). Plant grafting: new mechanisms, evolutionary implications. *Frontiers in Plant Science*, 5.
- Gong, X., Shi, S., Dou, F., Song, Y., & Ma, F. (2017). Exogenous melatonin alleviates alkaline stress in *malus hupehensis* rehd. by regulating the biosynthesis of polyamines. *Molecules*, 22(9), 1542.
- Güney, M., Aydöner Çoban G., & Güney, M. (2024, November 1). Chitosan nanoparticles and their influence on gene expression and secondary metabolism in fruits. *Balkan 12th International Conference on Applied Sciences*.
- Güney, M., Kafkas, S., Keles, H., Aras, S., & Ercişli, S. (2018). Characterization of hawthorn (*Crataegus* spp.) genotypes by SSR markers. *Physiology and molecular biology of plants*, 24(6), 1221-1230.
- Güney, M., Ergun, Z., & Gündeşli, M. A. (2022). The variation of fatty acid composition between fresh and stored avocado (*Persea americana*). *International Journal of Agriculture Forestry and Life Sciences*, 6(2), 51-54.
- Güney, M., Kafkas, S., Keles, H., Zarifikhosroshahi, M., Gündesli, M. A., Ercişli, S., ... & Bujdoso, G. (2021). Genetic diversity among some walnut (*Juglans regia* L.) genotypes by SSR markers. *Sustainability*, 13(12), 6830.
- Güney, M., Özyalın, S., & Keleş, H. (2024, November 8–10). Analysis of plant stress response mechanisms using RNA sequencing. *17th International Black Sea Conference on Applied Sciences*, Rize, Türkiye, 198–210.
- Güney, M., Özyalın, S., & Keleş, H. (2024, November 8–10). Gene expression and metabolic pathways in fruit quality: Flavor, color, and nutritional value. *Karadeniz 17th International Conference on Applied Sciences*, Rize, Türkiye, 159–168.
- Han, Y., Beck, W., Mewis, I., Förster, N., & Ulrichs, C. (2023). Effect of ozone stresses on growth and secondary plant metabolism of *Brassica campestris* l. ssp. *chinensis*. *Horticulturae*, 9(9), 966.
- Hao, X., Ren, J., Xu, M., Sun, B., Li, R., Yang, S., ... & Xu, W. (2025). Melatonin in plant pathogen defense: a review of its role in horticultural crops. *Horticulture Research*, 12(9).
- Hassan, H., Mohamed, A., Feleafel, M., Salem, M., Ali, H., Akrami, M., ... & Abd-Elkader, D. (2021). Natural plant extracts and microbial antagonists to control fungal pathogens and improve the productivity of zucchini (*Cucurbita pepo* L.) in vitro and in greenhouse. *Horticulturae*, 7(11), 470.
- Hernández-Hierro, J. M., Cozzolino, D., Feng, C. H., Rato, A. E., & Nogales-Bueno, J. (2022). Recent Advances of Near Infrared Applications in Fruits and Byproducts. *Frontiers in Plant Science*, 13, 858040.

- Hou, Y., Zhai, L., Li, X., Xue, Y., Wang, J., Yang, P., ... & Bian, S. (2017). Comparative analysis of fruit ripening-related miRNAs and their targets in blueberry using small rna and degradome sequencing. *International Journal of Molecular Sciences*, 18(12), 2767.
- Hu, S., Wang, T., Shao, Z., Meng, F., Chen, H., Wang, Q., ... & Liu, L. (2022). Brassinosteroid biosynthetic gene *slcyp90b3* alleviates chilling injury of tomato (*Solanum lycopersicum*) fruits during cold storage. *Antioxidants*, 11(1), 115.
- Jain, S., Kore, D., Kishorkumar, G., Mohapatra, A., Baksh, H., Kumar, V., ... & Haokip, S. (2023). A comprehensive review on protected cultivation of horticultural crops: present status and future prospects. *International Journal of Environment and Climate Change*, 13(11), 3521-3531. <https://doi.org/10.9734/ijecc/2023/v13i113528>
- Juxian, G., TingQuan, W., Mei, F., Li, G., Luo, W., Yunyan, K., ... & Wang, T. (2022). TCP15 negatively regulates anthocyanin biosynthesis under low light in *Brassica*. (Preprint).
- Kang, H., Thomas, H., Xia, X., Shi, H., Zhang, L., Hong, J., ... & Zhou, Y. (2025). An integrative overview of cold response and regulatory pathways in horticultural crops. *Journal of Integrative Plant Biology*, 67(4), 1028-1059.
- Kim, M., Yu, W., Song, D., Chun, S., Kim, M., Lee, A., ... & Mo, C. (2024). Prediction of soluble-solid content in citrus fruit using visible–near-infrared hyperspectral imaging based on effective-wavelength selection algorithm. *Sensors*, 24(5), 1512.
- Ku, H. and Ha, S. (2020). Improving nutritional and functional quality by genome editing of crops: status and perspectives. *Frontiers in Plant Science*, 11.
- Kumari, A., Singh, S., Mathpal, B., Verma, K., Garg, V., Bhattacharyya, M., ... & Bhatt, R. (2023). The biosynthesis, mechanism of action, and physiological functions of melatonin in horticultural plants: a review. *Horticulturae*, 9(8), 913.
- Kusumiyati, K., Hadiwijaya, Y., & Putri, I. (2019). Non-destructive classification of fruits based on vis-nir spectroscopy and principal component analysis. *Jurnal Biodjati*, 4(1), 89-95.
- Li, Q., Sapkota, M., & Knaap, E. (2020). Perspectives of CRISPR/CAS-mediated cis-engineering in horticulture: unlocking the neglected potential for crop improvement. *Horticulture Research*, 7(1).
- Li, R. and Cheng, Y. (2023). Recent advances in mechanisms underlying defense responses of horticultural crops to *Botrytis cinerea*. *Horticulturae*, 9(11), 1178.

- Macias-González, M., Truco, M., Han, R., Jenni, S., & Michelmore, R. (2021). High-resolution genetic dissection of the major qtl for tipburn resistance in lettuce, *Lactuca sativa*. *G3 Genes|genome|genetics*, 11(7).
- Özyalın, S., & Yaman, C. (2023). Tıbbi Adaçayı (*Salvia officinalis*)'nın İn vitro Çoğaltımı Üzerine Temel Besin Ortamlarının ve Büyüme Düzenleyici Tiplerinin Etkisi. *Kahramanmaraş Sütçü İmam Üniversitesi Tarım ve Doğa Dergisi*, 26(3), 600-609.
- Özyalın, S. (2023). Şeftalide hasat ve depolama. In A. Endes & S. Aras (Eds.), *Şeftali Yetiştiriciliği, Hastalık ve Zararlıları* (pp. 107-124). Ankara: İK-SAD Publishing House.
- Parmar, N., Singh, K., Sharma, D., Singh, L., Kumar, P., Nanjundan, J., ... & Thakur, A. (2017). Genetic engineering strategies for biotic and abiotic stress tolerance and quality enhancement in horticultural crops: a comprehensive review. *3 Biotech*, 7(4).
- Perato, S., Furio, R., Tomas-Grau, R., Pilar, M., Hael-Conrad, V., Ricci, J., ... & Martínez-Zamora, M. (2020). The fungal elicitorases require a functional ethylene pathway to activate the innate immunity in strawberry. *Plant Biology*, 22(6), 1030-1040.
- Pitt, H. (2021). What knowledge is required to grow food? a framework for understanding horticulture's skills 'crisis'. *Journal of Rural Studies*, 85, 59-67.
- Priyanka, K., Joshi, S., Bisen, M., Sumit, S., Singh, D., Nayak, R., ... & Siddiqua, A. (2025). Crispr-cas9 applications in horticultural crop improvement. *Plant Cell Biotechnology and Molecular Biology*, 26(1-2), 133-141.
- Roy, A., Chaurasia, H., Kumar, B., Kumari, N., Jaiswal, S., Srivast, M., ... & Kumar, D. (2024). Featl: a comprehensive web-based expression atlas for functional genomics in tropical and subtropical fruit crops.
- Sen, H., Kumar, A., & Janeja, H. (2024). Biofortification of major crops through conventional and modern biotechnological approaches to fight hidden hunger: an overview. *Journal of Advances in Biology & Biotechnology*, 27(7), 96-113.
- Shamshin, I., Dubrovsky, M., Трифонова, А., Борис, К., & Kudryavtsev, A. (2023). Powdery mildew resistance of apple clonal rootstocks from the collection of the Michurinsk State Agrarian University. *Vavilov Journal of Genetics and Breeding*, 27(6), 572-581.
- Sharma, A., Kumar, S., Singh, A., Kumar, S., Yadav, H., Hazarika, S., ... & Hasan, M. (2024). Exploring the role of robotic automation in climate vulnerability mitigation: towards sustainable horticulture. *International Journal of Environment and Climate Change*, 14(2), 6-13.

- Shibuya, T., Itagaki, K., Tojo, M., Endo, R., & Kitaya, Y. (2011). Fluorescent illumination with high red-to-far-red ratio improves resistance of cucumber seedlings to powdery mildew. *Hortscience*, 46(3), 429-431.
- Su, D., Wu, M., Wang, H., Shu, P., Song, H., Yu, S., ... & Deng, H. (2025). Bi-functional transcription factor SLBHLH95 regulates fruits flavonoid metabolism and grey mould resistance in tomato. *Plant Biotechnology Journal*, 23(6), 2083-2094.
- Sun, C., Zhang, Y., Liu, L., Liu, X., Li, B., Jin, C., ... & Lin, X. (2021). Molecular functions of nitric oxide and its potential applications in horticultural crops. *Horticulture Research*, 8(1).
- Tejasree, A., Mirza, A., & Joka, V. (2024). Deciphering nature's secret of seaweed extract as a biostimulant on horticultural crops: a review. *Journal of Experimental Agriculture International*, 46(6), 417-427.
- Terefe, M. (2018). Biosafety issues of genetically modified crops: addressing the potential risks and the status of GMO crops in Ethiopia. *Cloning & Transgenesis*, 07(02).
- Tiwari, R., Lal, M., Kumar, R., Chourasia, K., Naga, K., Kumar, D., ... & Zinta, G. (2021). Mechanistic insights on melatonin-mediated drought stress mitigation in plants. *Physiologia Plantarum*, 172(2), 1212-1226.
- Tyagi, S., Kumar, R., Kumar, V., Won, S. Y., & Shukla, P. (2021). Engineering disease resistant plants through CRISPR-Cas9 technology. *GM crops & food*, 12(1), 125-144.
- Wang, H., Wang, H., Shao, H., & Tang, X. (2016). Recent advances in utilizing transcription factors to improve plant abiotic stress tolerance by transgenic technology. *Frontiers in Plant Science*, 7.
- Wang, R., Shu, P., Zhang, C., Zhang, J., Chen, Y., Zhang, Y., ... & Liu, M. (2021). Integrative analyses of metabolome and genome-wide transcriptome reveal the regulatory network governing flavor formation in kiwifruit (*Actinidia chinensis*). *New Phytologist*, 233(1), 373-389. <https://doi.org/10.1111/nph.17618>
- Wang, W., Wang, P., Li, X., Wang, Y., Tian, S., & Qin, G. (2021). The transcription factor slhy5 regulates the ripening of tomato fruit at both the transcriptional and translational levels. *Horticulture Research*, 8(1).
- Wang, Y., Jiang, H., Mao, Z., Liu, W., Jiang, S., Xu, H., ... & Chen, X. (2021). Ethylene increases the cold tolerance of apple via the MDERF1B-MD-CIBHLH1 regulatory module. *The Plant Journal*, 106(2), 379-393.
- Wei, L., Zhong, Y., Wu, X., Wei, S., & Liu, Y. (2024). Roles of nitric oxide and brassinosteroid in improving fruit quality during postharvest: potential regulators?. *Journal of Agricultural and Food Chemistry*, 72(43), 23671-23688.



- Xu, H., Li, M., Ma, D., Gao, J., Tao, J., & Meng, J. (2024). Identification of key genes for triacylglycerol biosynthesis and storage in herbaceous peony (*Paeonia lactiflora* pall.) seeds based on full-length transcriptome. BMC Genomics, 25(1).
- Xu, Y., Hu, W., Song, S., Ye, X., Ding, Z., Liu, J., ... & Jin, Z. (2023). MaDREB1F confers cold and drought stress resistance through common regulation of hormone synthesis and protectant metabolite contents in banana. Horticulture Research, 10(2), uhac275.
- Yin, L., Wang, P., Li, M., Ke, X., Li, C., Liang, D., ... & Ma, F. (2013). Exogenous melatonin improves *Malus* resistance to Marssonina apple blotch. Journal of Pineal Research, 54(4), 426-434. <https://doi.org/10.1111/jpi.12038>
- Zeng, Y., Pan, Z., Ding, Y., Zhu, A., Cao, H., Xu, Q., ... & Deng, X. (2011). A proteomic analysis of the chromoplasts isolated from sweet orange fruits [*Citrus sinensis* (L.) osbeck]. Journal of Experimental Botany, 62(15), 5297-5309.
- Zhang, A., Zhang, S., Wang, F., Meng, X., Ma, Y., Guan, J., ... & Zhang, F. (2023). The roles of microRNAs in horticultural plant disease resistance. Frontiers in Genetics, 14.
- Zhang, R., Zhang, C., Yu, C., Dong, J., & Hu, J. (2022). Integration of multi-omics technologies for crop improvement: status and prospects. Frontiers in Bioinformatics, 2.
- Zhang, S., Zheng, X., Reiter, R., Feng, S., Wang, Y., Liu, S., ... & Ren, M. (2017). Melatonin attenuates potato late blight by disrupting cell growth, stress tolerance, fungicide susceptibility and homeostasis of gene expression in *Phytophthora infestans*. Frontiers in Plant Science, 8.
- Zhang, X., Francis, M., Dawson, W., Graham, J., Orbović, V., Triplett, E., ... & Mou, Z. (2010). Over-expression of the *Arabidopsis* NPR1 gene in citrus increases resistance to citrus canker. European Journal of Plant Pathology, 128(1), 91-100.
- Zhang, Z., Chen, Z., Song, H., & Cheng, S. (2023). From plant survival to thriving: exploring the miracle of brassinosteroids for boosting abiotic stress resilience in horticultural crops. Frontiers in Plant Science, 14.
- Zhao, H., Zhang, T., Meng, X., Song, J., Zhang, C., & Gao, P. (2023). Genetic mapping and QTL analysis of fruit traits in melon (*Cucumis melo* L.). Current Issues in Molecular Biology, 45(4), 3419-3433.
- Zheng, X., Giuliano, G., & Al-Babili, S. (2020). Carotenoid biofortification in crop plants: citius, altius, fortius. Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids, 1865(11), 158664.

- Zhou, R., Jiang, F., Niu, L., Song, X., Yu, L., Yang, Y., ... & Wu, Z. (2022). Increase crop resilience to heat stress using omic strategies. *Frontiers in Plant Science*, 13.
- Zhu, H., Zhang, Y., Tang, R., Qu, H., Duan, X., & Jiang, Y. (2019). Banana transcriptome and degradome identify microRNAs functioning in differential responses to temperature stress. *BMC Genomics*, 20(1).